Harmonisation Efforts in AML MRD

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Sylvie Freeman DISCLOSURES OF COMMERCIAL SUPPORT

Name of Company	Research support	Employee	Consultant	Stockholder	Speaker's Bureau	Scientific Advisory Board	Other
Novartis	No	No	No	No	Yes	Yes	
Jazz	Yes	No	No	No	Yes	No	
BMS	Yes	No	No	No	No	No	

Flow cytometric MRD in AML is a high risk test (IVDR C)

Intended Purpose

- To monitor response to treatment
- As an aid to treatment choices
 - intensification and transplant related decisions time-points post induction, pre-transplant, post-transplant
 - more recently enrollment criterion for some trials



population and intended user

Adapted from from MedTech Europe 2023 IVDR ebook

Published Evidence Source





Expert Consensus Recommendations

2021 Update on MRD in acute myeloid leukemia: a consensus document from the European LeukemiaNet MRD Working Party

Michael Heuser,¹ Sylvie D. Freeman,² Gert J. Ossenkoppele,³ Francesco Buccisano,⁴ Christopher S. Hourigan,⁵ Lok Lam Ngai,³ Jesse M. Tettero,³ Costa Bachas,³ Constance Baer,⁶ Marie-Christine Béné,⁷ Veit Bücklein,⁸ Anna Czyz,⁹ Barbara Denys,¹⁰ Richard Dillon,¹¹ Michaela Feuring-Buske,¹² Monica L. Guzman,¹³ Torsten Haferlach,⁶ Lina Han,¹⁴ Julia K. Herzig,¹² Jeffrey L. Jorgensen,¹⁵ Wolfgang Kem,⁶ Marina Y. Konopleva,¹⁴ Francis Lacombe,¹⁶ Marta Libura,¹⁷ Agata Majchrzak,¹⁸ Luca Maurillo,⁴ Yishai Ofran,¹⁹ Jan Philippe,¹⁰ Adriana Plesa,²⁰ Claude Preudhomme,²¹ Farhad Ravandi,¹⁴ Christophe Roumier,²¹ Marion Subklewe,⁸ Felicitas Thol,¹ Arjan A. van de Loosdrecht,³ Bert A. van der Reijden,²² Adriano Venditti,⁴ Agnieszka Wierzbowska,²³ Peter J. M. Valk,²⁴ Brent L. Wood,²⁵ Roland B. Walter,²⁶ Christian Thiede,^{27,28} Konstanze Döhner,¹² Gail J. Roboz,¹³ and Jacqueline Cloos³

Technical Aspects of Flow Cytometry-based Measurable Residual Disease Quantification in Acute Myeloid Leukemia: Experience of the European LeukemiaNet MRD Working Party

Jesse M. Tettero¹, Sylvie Freeman², Veit Buecklein³, Adriano Venditti⁴, Luca Maurillo⁴, Wolfgang Kern⁵, Roland B. Walter⁶, Brent L. Wood⁷, Christophe Roumier⁸, Jan Philippé⁹, Barbara Denys⁹, Jeffrey L. Jorgensen¹⁰, Marie C. Bene¹¹, Francis Lacombe¹², Adriana Plesa¹³, Monica L. Guzman¹⁴, Agnieszka Wierzbowska¹⁵, Anna Czyz¹⁶, Lok Lam Ngai¹, Adrian Schwarzer¹⁷, Costa Bachas¹, Jacqueline Cloos¹, Marion Subklewe³, Michaela Fuering-Buske¹⁸, Francesco Buccisano⁴

Harmonisation as an aid to evidence of Analytical and Clinical Performance



Cytometer and settings

Antibody Panels

2021 ELN Recommended core combination

ELN Tube PerCP-Alexa PE FITC PECy7 APC PB V500 KO Cy5.5 Fluor750 Fluorochorme **CD34** CD13 CD7 CD33 **CD56** CD117 HLA-DR CD45 Antigen 8G12 L138 M-T701 D3HL60.251 NCAM16.2 104D2D1 HI30 Clone Immu357 J33 Manufacturer ΒD ΒD ΒD BC BC BD BC BC ΒD Order No. 345801 347406 561602 B92408 341027 B92450 B36291 560777 B36294 5 µl 5 µl 5 µl 5μl 5 µl 5 µl 5 µl 5 µl 5 µl Amount **Hovon P1 Tube** PerCP-APC-H7 PE PECy7 FITC APC **BV421** KO Cy5.5 Fluorochorme CD7 **CD56 CD34** CD117 **CD33** HLA-DR CD13 CD45 Antigen M-T701 MY31 8G12 104D2D1 P67.6 L243 (G46-6) **WM15** J33 Clone Manufacturer ΒD ΒD ΒD BC ΒD ΒD BD BC 347222 Order No. 555360 345810 B49221 345800 641411 562596 B36294 5 ul 5 µl 5 µl 5μl 5 µl 5 µl 5 ul Amount 5 µl

CD34 CD117 CD33 CD13 HLADR CD45 CD7 CD56

Also highly relevant

- Myeloid maturation / monocytic markers
- CD38 with CD34 and 'LSC' markers (markers absent on normal CD34+CD38low/neg cells)

Antibody Panels

Progress in technology

- More colours with new cytometers
- More / new markers

ORIGINALARTICLE

CLINICAL CYTOMETRY

Validation of a 12-color flow cytometry assay for acutemyeloid leukemia minimal/measurable residual diseasedetectionWang et alAugust 2023

gene expression array 28 markers but identified <1% further MRD+ CD9 CD52 CD44 0 °0 0 MFI (x 10³) 8000 <u>مە</u>م 000 <u>00</u> 00 0000 0 MRD D MRD MRD D D 151 0.01 0.1 1 10 100 CD54 CD97 CD300a/c 321 201 % MRD by standard markers 0

MFI (x 10³)

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MRD

Adapted from Coustan-Smith et al. JCI Insight 2018

plasma membrane proteomics

overlap in identified aberrant markers



Adapted from de Boer, et al. Cancer Cell 2018

Antibody Panels

AML MRD panel composition survey: 11 ELN Laboratories
 Belgium (1), France (2), Germany (2), Israel (1), Netherlands (1), Poland (1), UK (1), USA (2)

Excluding 8c ELN core combination - 29 markers in use.

Asynchronous	Lymphoid	LSC-specific	Monocytic	Misc
CD38	CD19	CD38	CD64	CD123
CD133	CD22	CD45RA	CD14	CD36
	CD2	CLL-1/CLEC12A	CD300e	CD54
CD11b	CD5	TIM3		CD25
CD15	CD9	CD90		CD71
CD65		CD97		
		GPR56		
		CD81		
		CD97		
		CD200		
		CD44		

Work in progress:

- Updating Consensus markers for:
 - 10c tube
 - 12c tube
 - 13-16c tube
 - LSC specific
 - Monocytic specific

Analytical performance

Use of proficiency testing material to harmonize results

- 1. Sample exchange schemes
- 2. External quality assurance schemes
 - UK NEQAS Leukocyte immunophenotyping



UK NEQAS

Leucocyte Immunophenotyping

F	Results and Performance	_							
	Percentage MRD Population	Your Res (%)	ults	Robust M (%)	lean	Robus (%	t SD)		
		0.049	0	0.2000)	0.15	82		
	Percentage MRD Population	z Score*	Performance Status		Performance Status Classifie			ation Over	12 Sample Period
			for th	lis Sample	Sati	isfactory	Act	tion	Critical
[12	(0	0

*z Score Limits Definitions

Please note the scale below is applicable to the tables above and to the z score histograms and Shewhart control charts that follow. It is <u>not</u> applicable to the Cusum control charts.



Histograms of Participant z Scores



98 laboratories participating in current UK NEQAS programme

- Uses stabilised EQA material with 4 distributions per year
- Test for panel, processing and analysis
- MRD samples now also contain 'normal' progenitors (PB stem cells)

UK NEQAS Leucocyte Immunophenotyping

Matthew Fletcher, Liam Whitby, Stuart Scott

August 2023 Exercise

Robust Mean	Robust SD
(%)	(%)
0.0914	0.0300

ELN MRD threshold 0.1%

Robust SD - 0.07% to 0.12%

Understanding Obstacles for harmonisation

EQAs are a tool to inform issues and guide changes in practice

ELN-NEQAS Interlaboratory study for molecular MRD

- *NPM1^{mut}* PCR MRD -High proportion of false positive results in EQA negative sample
 - Proportion attributed to use of a specific highly active reverse transcriptase

– more likely to introduce errors (less of an issue for rearranged genes). Scott et al Blood Advances 2023

Issues for Flow Cytometric MRD Panels vs Processing vs Analysis & Interpretation

Analytical performance – development of an electronic EQA NEQAS /ELN

EQA pilot - Flow files (FCS) of diagnostic, control and MRD samples stained with ELN tube

Items for harmonization

- Denominator used to calculate the MRD population size
- ELN Marker Staining Intensity of Diagnostic Blasts
- Presence and Ranking of Diagnostic LAIP(s)
- What is the Aberrant Immunophenoype quantified in the MRD sample? Diagnostic or emerging Different from normal
- MRD value (% and events) in MRD sample
 & Normal Control (ie background)
- LOQ by events criteria (to assess sample quality)



Matthew Fletcher Liam Whitby Stuart Scott



Lessons learnt from initial pilots:

Confounder for harmonized values

CD34+ vs CD117+ for reporting MRD % particularly if dominant aberrant IP changes 34 or 117 expression between diagnosis and MRD

Analytical performance – development of an electronic EQA

MRD Population (%)	z score	Performance Status by z Score
CD34+	-0.405	Satisfactory
CD117+	-0.199	Satisfactory

z-score for MRD %, but also performance by reference value



Diagnostic LAIP blasts

In silico dilution

→ MRD sample with n LAIP cells n = reference / maximum MRD events



Participant-specific Report

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ocyte Immu	inophenot	typing								NHS Foundati	on Trust
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iagnostic LAI	P on CD117	7+ Blasts	0055-1-1		2022-14 (212.11				
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Satisfactory

Satisfactory

CD34+

CD117+

Myeloma / CLL / ALL MRD LOD 20-30 cells LOQ 50 cells

Absolute LOQ - allows harmonization of sample quality assessment for AML MRD Critical to qualify a negative result

Sample LOQ of 0.01% if 500,000 leukocytes acquired

Next steps to consider

- blast "LOQ" need 500 blast events to detect MRD target present in 1 in 10 blasts
- assessment of hemodilution particularly if MRD < 0.1%

Use of absolute Limit of Quantification in AML MRD



Improving Flow cytometric MRD prognostic accuracy - Clinical Performance



F. Buccisano, personal communication & *Buccisano et al Haematologica. 2022*

J Tettero et al EHA 2023 <u>Hemasphere.</u> 2023 Aug; 7(Suppl)

Analysis strategies: considerations for performance and harmonization

LAIP - leukemia immunophenotypic footprint

- number of Boolean gates and exact gate positions varies between SOPs, operators and samples
- can achieve MRD^{negative} < absolute LOQ with multiple Boolean gates + empty space footprint

Different from Normal

- applicable to: immunophenotypic shifts, when no diagnostic sample & longer-term monitoring
- screens several plots of 2 marker combinations, usually focused on blast compartments
- can be standardized by fixed gates for the most useful empty spaces
 - allows rapid analysis
- may have higher background from normal controls than multiple Boolean gates

Flow cytometric MRD target assessment needs control bone marrows



Leukemic Aberrant ImmunoPhenotype LAIP = abnormal immunophenotype of diagnostic leukaemic blasts

Sensitive / specific LAIPs for tracking MRD amongst normal regenerating blasts are in immunophenotypic 'empty spaces' of normal blasts

Control BMs required to interpret positive result vs background

Different from Normal DfN analysis relies on these empty spaces defined by set panel
 Control BMs required to define empty spaces and their background

Empty spaces? - Consider "aberrant immunophenotypes with indeterminate leukemic potential"

Aberrant immunophenotypes with indeterminate leukemic potential



Impacts: Reporting, scientific validity and clinical performance

• Library of reference 'control' bone marrows - need sufficient number and subtypes



Other Lab Control BM CD34+



Aberrant immunophenotypes with indeterminate leukemic potential

Also observed for aberrant lymphoid expression

80 patients 80 70 60 Normal Distribution (%) Atypical 50 Abnormal 40 30 20 10 CD13/ CD13/ CD2 CD7 **CD56** HLA-DR **CD16** Flow Abnormality On CD34+ blasts

Immunophenotypic abnormalities in CCUS

Jevremovic et al, AJCP 2022



W Kern et al, Cytometry Part B Clinical Cytometry 2023

Range of control BMs impacts MRD result specificity - especially results < 0.1%



G Gui NIH USA / F Dumezy, C Roumier, A Plesa ALFA France

Conclusions

Harmonisation leading to equivalent MRD results between laboratories - key step for Clinical Evidence

- Processing including stable cytometer settings newer cytometers
- Development of an analytical EQA with reference value to monitor performance & competency Includes: identification and ranking of common useful LAIPs /DfN for AML MRD MRD % from 1) CD34+ blasts 2) CD117+ blasts
 Integration of control sample for % background by LAIP /DfN gating
- Sample quality assessment absolute LOQ / blast events / hemodilution
- Range of control bone marrows to exclude false positives from "aberrant immunophenotypes with indeterminate potential"
- LOQ or <0.01% may differentiate deeper responders in future